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Description

The present invention relates to vaccines against Bordetella pertussis, Bordetella parapertussis or Bordetella bronchiseptica.

Bordetella pertussis is the causative agent of whooping cough in man, although outbreaks of the disease have been associated with the related organism Bordetella parapertussis. Bordetella bronchiseptica is primarily an animal pathogen, although it has been isolated from children with whooping cough-like symptoms. Immunization programmes with whole-cell vaccine of B. pertussis have been relatively effective in controlling the disease, although currently the uptake of the vaccine is low in some developed countries because of the reactogenicity associated with vaccination. Clinical symptoms noted for the 1 in 10,000 children suffering side-effects may include persistent screaming, fever and local reactions.

There is a need for a new pertussis acellular vaccine that lacks the components present in the whole cell vaccine associated with reactogenicity, but still comprises the protective epitopes. The search for the protective components has centered on a number of outer-membrane associated antigens. These include pertussis toxin (ptx Lymphocytosis promoting factor/LPF), filamentous haemagglutinin (FHA), cytotoxic adenylate cyclase (Adcase), dermonecrotic toxin (DNT), tracheal cytotoxin, the agglutinogens (Agg2, Agg3), the 69kDa outer membrane protein (omp) (P.69) and lipopolysaccharide (LPS).

Much work has been carried out on the pertussis toxin (LPF), which is believed by many to be the most important part of any acellular pertussis vaccine (Bacterial Vaccines, 1984, Chapter 3, Manclark *et al.* Ed: Germainer). The results of a recent clinical trial in Sweden with an LPF/FHA vaccine showed that such a vaccine provided only about 69% protection (Lancet 1, 995, 1988). This is lower than the results expected for protection provided by a whole-cell vaccine, and resulted in the LPF/FHA vaccine not being licensed by the Swedish Health Authority.

EP-A-0162639 discloses a vaccine formulation for protection against B. pertussis which includes an antigenic preparation derived from B. pertussis comprising proteinaceous material associated with adenylate cyclase activity (ACAP) together with a pharmaceutically acceptable carrier therefor. An ACAP with a molecular weight of 69kDa is disclosed.

In order to characterise which regions of this ACAP, termed the P.69 antigen, were important for protection the gene encoding the protein was cloned and sequenced (Charles, *et al.* PNAS, Vol 86, 3554-3558, 1989). P.69 as extracted from B. pertussis appears to be the processed form of a 93kDa precursor (P.93).

The present invention provides a polypeptide suitable for use in a vaccine which polypeptide presents an epitope comprising:

- (a) the amino acid sequence coded for by nucleotides CCGGGTCCCCAGCCGCCG which are nucleotides 1885 to 1902 of the P.69 gene of B. pertussis CN2992; or
 - (b) the corresponding amino acid sequence of another strain of B. pertussis or of a strain of B. parapertussis or B. bronchiseptica;
- the said polypeptide being no more than 50 amino acid residues long or being a chimaeric protein having an amino acid sequence comprising the sequence of a carrier protein and a foreign sequence of no more than 50 amino acid residues which comprises the sequence of a said epitope.

The polypeptides of the invention comprise a defined antigenically effective sequence. This sequence is

PGPQPP

based on the sequence of the P.69 gene of B. pertussis CN2992 disclosed by Charles *et al.* (1989) and using the one letter code for amino acids (Eur. J. Biochem. 138, 9-37, 1984). The sequence therefore essentially consists of amino acid residues 547 to 552 of the P.69 protein of B. pertussis CN2992. The corresponding sequence for other strains of B. pertussis and for strains of B. parapertussis and B. bronchiseptica can be readily determined by lining up the amino acid sequence of the P.69 antigen, the P.70 antigen or the P.68 antigen respectively with the P.69 sequence shown by Charles *et al.* (1989).

Preferably the epitope comprises:

- (a₁) the amino acid sequence coded for by nucleotides

CGCGCCGAGCCGGGTCCCCAGCCGCCGAGCCGCCGAGCCGCCGAGC

CGGAAGCGCCGGCGCCGCAACCG

(b.) the corresponding amino acid sequence of another strain of B. pertussis or of a strain of B. parapertussis or B. bronchiseptica;

Sequence (a_1) is therefore:

5 APQPGPQPPQPPQPQPEAPAPQP

The sequence represents amino acid residues 544 to 566 of the P.69 protein of *B. pertussis* CN2992. This sequence and the corresponding sequence (b₁) for the P.70 antigen of *B. parapertussis* and the P.68 antigen of *B. bronchiseptica* can be aligned as follows. The PGPQPP epitope is underlined:

P.70 APQPGPQGPPQP

10 P.68 APQPGPQPGPQPPQPPQPPQP-----PQRQPEAPAPQP

P.69 APQPGPQP-PQPPQP-----QPEAPAPQP

A first type of polypeptide according to the invention is composed of up to 50 amino acid residues, for example up to 40 or up to 30 or up to 20 amino acid residues, which presents an epitope as defined above. Further amino acids may therefore be added to one or both ends of the epitope. One, two, three or four additional residues may be provided at the N-terminus or C-terminus or at both termini of the defined epitope.

Where additional residues are provided at either or both ends of the epitope, preferably these are the natural residues. These can be deduced from the sequence of the P.69 antigen of *B. pertussis*, of the P.70 antigen of *B. parapertussis* or of the P.68 antigen of *B. bronchiseptica*. Preferred flanking sequences for the epitope are therefore the flanking sequences which naturally occur either side of the amino acid sequence for the epitope in the overall sequence of the P.69, P.70 or P.68 protein as the case may be. Also, a cysteine residue may be provided at the N-terminus or C-terminus. In particular, a cysteine residue may be added to the C-terminus alone. This is in order to facilitate carrier coupling and/or to enhance the immunogenicity of the polypeptide.

The polypeptide may have a free C-terminal carboxy group. Alternatively, it may be in the form of a C-terminal amide. Pharmaceutically acceptable salts of the polypeptide may be employed. The polypeptide may be coupled to a carrier in order to create an immunogen which is antigenically active. Any appropriate physiologically acceptable carrier may be employed. A conjugate between the polypeptide and the carrier may be formed. The carrier may be for example bovine serum albumin, thyroglobulin, ovalbumin, keyhole limpet hemocyanin (KLH) or hepatitis B core antigen.

A second type of polypeptide according to the invention is a chimaeric protein which presents the defined epitope. The chimaeric protein is typically a carrier protein which has been modified so that its amino acid sequence comprises a foreign sequence of up to 50 amino acids which includes the sequence of the desired epitope. Some amino acids of a protein may be replaced by the foreign amino acid sequence. Alternatively, the foreign amino acid sequence is fused to a protein. An intervening linker of up to 10 amino acids, for example of up to 5 amino acids, may be provided between the epitope and the carrier. The foreign amino acid sequence may vary in length as described for the first type of polypeptide according to the invention.

The epitope is exposed on the surface of the chimaeric protein so that it is presented to the immune system. The chimaeric protein may take the form of a particle or form part of a particulate aggregation. Such an aggregation may comprise plurality of chimaeric proteins and/or may be a viral particle. A protein to which a foreign amino acid sequence comprising the epitope may be fused may be a particle-forming protein such as hepatitis B surface antigen (HBsAg, Ep-A-0175261) or hepatitis B core antigen (HBcAg, JP-A-63196299). The foreign sequence may be inserted into the sequence of a viral protein exposed on the surface of the virus (GB-A-2125065). The viral protein may be a capsid protein of a virus.

The foreign sequence may therefore be provided at one of the antigenic sites of a picornavirus such as poliovirus (EP-A-0302801). The epitope may be presented at one of the antigenic sites, for example site 1, 2 or 3, on a capsid protein of an attenuated strain of type 1 poliovirus, or at an antigenic site of type 2 or 3 poliovirus. Other picornaviruses, suitably modified, may be used, e.g. Bovine enterovirus.

The amino acid sequence of an antigenic site of a picornavirus may be replaced completely or partly by the foreign amino acid sequence. Preferably the foreign amino acid sequence is provided in place of some or all of antigenic site 1 of an attenuated strain of type 1 poliovirus. The attenuated strain is typically the Sabin 1 vaccine strain. Antigenic site 1 of a type 1 poliovirus is composed of amino acid residues 91 to 102 of the VP1 capsid protein.

The polypeptides of the invention are synthetic polypeptides. They may be prepared by chemical synthesis, in particular the first type of polypeptide of up to 50 amino acid residues long. Solid-phase or solution methods of peptide synthesis may be employed. A polypeptide can be built up therefore by a process comprising condensing single amino acids and/or preformed peptides or two or more amino acids

in the order in which amino acids occur in a polypeptide of the invention. The polypeptide may be synthesised so as to possess a free C-terminal carboxy group or a C-terminal amide group. If desired, the polypeptide may be converted into a pharmaceutically acceptable salt.

In solid-phase synthesis, the amino acid sequence of the desired polypeptide is built up sequentially from the C-terminal amino acid which is bound to an insoluble resin. When the desired polypeptide has been produced, it is cleaved from the resin. When solution-phase synthesis is employed, the polypeptide may again be built up from the C-terminal amino acid. The carboxy group of this acid remains blocked throughout by a suitable protecting group, which is removed at the end of the synthesis.

Whichever technique, solid phase or solution-phase, is employed each amino acid added to the reaction system typically has a protected α -amino group and an activated carboxy group. An amino group may be protected by the fluorenyl-methoxycarbonyl (Fmoc) or t-butoxycarbonyl (Boc) group. A carboxy group may be activated as a pentafluorophenyl or 1-oxo-2-hydroxy-dihydrobenzotriazine ester. Each condensation step may be effected in the presence of dicyclohexylcarbodiimide or 1-hydroxybenzotriazole.

Side chain functional groups are typically protected too, for example the side chain amino group of a lysine, the side chain hydroxy group of a threonine or the side chain sulphhydryl group of a cysteine. After each step in the synthesis, the α -amino protecting group is removed.

However, any side-chain protecting groups are generally only removed at the end of the synthesis although they may be retained if desired.

The polypeptides may be prepared with a C-terminal carboxy or amide group as desired. In solid phase peptide synthesis, this may be determined by how the C-terminal amino acid is linked to the resin support and/or how the final peptide is cleaved from the resin. Typically the resin is a styrene and/or divinylbenzene polymer. The C-terminal amino acid may be linked to the resin via an ester linkage which can be cleaved by a strong acid such as HBr in trifluoroacetic acid or HF to give the peptide with a C-terminal carboxy group. Ammonolysis can give the corresponding amide instead.

An alternative method of obtaining a polypeptide amide by solid phase synthesis is to arrange for the C-terminal amino acid of the polypeptide to be linked to the resin via a peptide aminobenzhydryl bond. This can be formed by coupling with dicyclohexylcarbodiimide and can be cleaved with HF, typically in the cold. For solution phase synthesis, whether a C-terminal carboxy or amide group is present may depend upon how the carboxy group of the C-terminal amino acid is blocked and, at the end of the synthesis, unblocked. A polypeptide with a C-terminal carboxy group can be converted into one with a C-terminal amide group and vice versa.

Both types of polypeptide according to the invention may be prepared by recombinant DNA methodologies, in particular by:

- (i) preparing an expression vector which incorporates a DNA sequence encoding the said polypeptide and which is capable of expressing the said polypeptide when provided in a suitable host; and
- (ii) providing the said vector in the said host such as to enable expression of the said polypeptide to occur.

Thus a DNA sequence encoding the desired polypeptide is provided. An expression vector is prepared which incorporates the DNA sequence and which is capable of expressing the polypeptide when provided in a suitable host. The DNA sequence is located between translation start and stop signals in the vector. Appropriate transcriptional and translational control elements are also provided, in particular a promoter for the DNA sequence and a transcriptional termination site. The DNA sequence is provided in the correct frame such as to enable expression of the polypeptide to occur in a host compatible with the vector.

In the case of a chimaeric protein, a DNA fragment encoding the foreign amino acid sequence is inserted into a vector at a location which enables the epitope of interest to be expressed, as part of the chimaeric protein, exposed on the surface of the protein. The chimaeric protein is then expressed. Cells harbouring the vector are cultured so as to enable expression to occur. Depending on the type of chimaeric protein, the protein may self-assemble into particles.

Any appropriate host-vector system may be employed. The vector may be plasmid. In that event, a bacterial or yeast host may be used for example *E. coli* or *S. cerevisiae*. Alternatively, the vector may be a viral vector. This may be used to transfect cells of a mammalian cell line, such as CHO cells, in order to cause polypeptide expression.

An epitope according to the invention may be linked to one or more helper T-cell (Th-cell) epitopes. A Th-cell epitope is a site capable of eliciting help for antibody production. A Th-cell epitope is capable of binding class II major histocompatibility (MHC) molecules on the surface of host antigen-presenting cells and B-cells subsequently interacting with the T-cell receptor in the form of a trimolecular complex in order to induce B-cells to differentiate and proliferate.

A Th-cell epitope may be linked to the first type of polypeptide of the invention in a variety of ways. Glutaraldehyde polymerisation may be used, in which the polypeptide of the invention is copolymerised with a polypeptide which presents a Th-cell epitope via their amino groups. The polypeptide of the invention and the polypeptide presenting the Th-cell epitope may be conjugated together via a heterobifunctional cross-linking agent such as m-maleimidobenzoyl-N-hydroxy-succinimide ester (MBS).

The polypeptide of the invention may alternatively be linked at its C-terminus or N-terminus to a polypeptide presenting a Th-cell epitope via a peptide bond. This may be achieved by co-linear synthesis of the polypeptide of the invention and the polypeptide presenting the Th-cell epitope or by use of recombinant DNA technology as above to express a fusion protein in which the two polypeptides are fused together. In any of the methods, any suitable Th-cell epitope may be used.

A preferred polypeptide presenting Th-cell epitopes is hepatitis B core antigen (HBcAg). A first type of polypeptide of the invention may be chemically coupled to HBcAg. Recombinant DNA technology can be used to produce a fusion protein according to the second type of polypeptide of the invention, comprising HBcAg to the amino terminus of which is linked the sequence of a polypeptide of the invention. The epitope of interest may be fused directly to the amino terminus of HBcAg. Alternatively, the sequence may be fused to the HBcAg via an intervening linker. Such a linker may be composed of one or more, for example up to ten, amino acid residues.

The polypeptides of the invention are useful as vaccines against B. pertussis, B. parapertussis or B. bronchiseptica. An effective amount of the polypeptide is administered to a host in need of vaccination. The polypeptide may be administered orally or parenterally, for example subcutaneously or intramuscularly. The polypeptide may be given in association with FHA. In this way, immunity to whooping cough may be induced in a human. Immunity to B. bronchiseptica may be induced in a mammal.

Typically, a polypeptide is administered orally or parenterally in an amount of 1 to 1000 µg per dose, more preferably from 10 to 100 µg per dose. A single dose may be given or a plurality of doses may be administered over a period of time. When FHA is administered, it may be used in an amount of 20 to 75 µg per dose. The FHA is typically administered in the same vaccine formulation as the polypeptide of the invention.

A vaccine of the invention comprises a polypeptide of the invention, optionally FHA, and a pharmaceutically acceptable carrier or diluent. The carrier or diluent may be any liquid medium suitable for use as a vehicle to introduce an antigen into a patient, for example an isotonic saline solution. The polypeptide may also be present with an adjuvant for stimulating the immune response and thereby enhancing the effect of the vaccine. A suitable physiologically acceptable adjuvant is aluminium hydroxide or aluminium phosphate.

Conveniently the vaccine formulations are presented to contain final concentration of antigenic polypeptide in a range of from 0.01 to 5mg/ml, preferably 0.03 to 2 mg/ml, most preferably about 0.3 mg/ml. After formulation the vaccine may be incorporated into a sterile container which is then sealed and stored at a low temperature, for example 4°C, or may be freeze-dried.

One or more doses of such a formulation may be administered to a host to be immunised. It is recommended that each dose is from 0.1 to 2 ml, preferably from 0.2 to 1ml and most preferably about 0.5ml of vaccine. Immunity can therefore be induced by the administration of an effective amount of a vaccine formulation.

The following Examples illustrate the present invention. In the accompanying drawings:

Figure 1 shows the construction of vector pWYG7.

Figure 2 shows the nucleotide sequence of the promoter region of GAL7. The synthesized promoter corresponds to the XhoI to BamHI fragment. Regions downstream of BamHI are present in native GAL7 including the RNA start site (↓) and the initiating ATG (underlined). The two basepairs which were altered to give a BamHI site are underlined.

Figure 3 shows the construction of pWYG7HBP.

Figure 4(a) shows the nucleotide sequence of synthetic oligo A containing the GAL7 untranslated leader sequence and the 5' region of the HBcAg gene.

Figure 4(b) shows the nucleotide sequence of synthetic oligo B, encoding the presumed BB05 epitope.

Figure 5 shows the results of Western blot of soluble proteins in Example 2(6) from induced yeast cells transformed with (1) pWYG7HBF (expressing FMD peptide-HBcAg fusion), (2) pWYG7HBF, (3) no plasmid and (4) pWYG7HBC (expressing HBcAg). The blot was developed using a rabbit anti-HBcAg serum and a goat anti-rabbit IgG-peroxidase conjugate. The position in the gel of 30K, 21.5K and 14.3KDa size markers is indicated.

Figure 6 shows the results of dot blot analysis in Example 2(7) of HBP fusion protein after sucrose density gradient centrifugation. Fraction 1 is the bottom fraction and fraction 20 the top. All the material reacting with the anti-HBcAg serum is in the middle fractions, indicating complete assembly of HBP into

particles.

Figure 7 shows the region of P.69 from which peptides 683, 684 and 685 and the epitope presented by the HBP fusion protein are derived. BB05/CORE binds BB05. Peptide 683 binds BB05, BB07, E4A8 and E4D7. Peptide 684: no binding. Peptide 685: binds PBE3. The numbering of the amino acid sequence refers to the residues of mature (i.e. signal sequence processed) P.69.

Figure 8 shows the analysis of peptides, covering residues 505 to 603 of the P.69 protein of *B. pertussis*, with monoclonal antibody.

EXAMPLE 1: Preparation of peptides

The peptides shown below were synthesised using an adaptation of the Merrifield method (Merrifield, JACS, 85, 2149-2154, 1963) described by Houghten (Houghten, PNAS, 82, 5131-5135, 1985):

Peptide 683: Δ PQPGPQPPQPPQPQPEAPAPQPPAGRELSC (peptide of invention)

Peptide 684: AGRELSAAANAANTGGVGLASTLWYAEC

Peptide 685: TLWYAESNALSKRLGELRLNPDAGGAWGRGC

Peptide 683 is composed of the amino acids coded for by nucleotides 1876 to 1962 of the P.69 gene and has an additional non-natural carboxy-terminal cysteine residue. Peptide 684 is composed of the amino acids coded for by nucleotides 1948 to 2031 of the P.69 gene and has an additional non-natural carboxy-terminal cysteine residue. Peptide 685 is composed of the amino acids coded for by nucleotides 2014 to 2100 of the P.69 gene and has an additional non-natural carboxy-terminal cysteine residue.

Each peptide was synthesised on a p-methyl -benzhydrylamine divinylbenzene resin. The α -amino protecting group on each amino acid was t-butoxycarbonyl (Boc). Each coupling cycle was as follows:

1. Wash resin with dichloromethane - 10 minutes

2. Wash with 5% diisopropylethylamine in dichloromethane - 2 minutes x 3

3. Dichloromethane wash - 1 minute x 2

4. Couple t-butoxycarbonyl amino acid in dichloromethane, 0.3M diisopropylcarbodiimide - 60 minutes. For N and Q coupling was effected in dimethylformamide, 0.3M diisopropylcarbodiimide and 0.125M hydroxybenzotriazole.

5. As 3

6. Deprotect with 50% trifluoroacetic acid in dichloromethane - 20 minutes

7. Dichloromethane wash - 1 minute x 6

8. Return to 2.

When coupling cycles were completed the peptide was cleaved off the resin using hydrogen fluoride for 1 hour with an anisole scavenger 10%. The peptide was thus obtained with a carboxy-terminal amide group. It was then ether washed, dried, dissolved in 15% acetic acid and lyophilized.

Example 2: Expression of *B. pertussis* epitope - HBcAg Fusion protein (HBP) in yeast

1. General

An amino acid sequence corresponding to nucleotides 1855 to 1944 of the P.69 gene was fused genetically to the amino-terminus of HBcAg. The resulting fusion protein was expressed efficiently in yeast and reacted with monoclonal antibody (Mab) BB05 which was raised against the 68kDa protein of *B. bronchiseptica* but cross-reacts with P.69 of *B. pertussis*. The fusion protein was shown to assemble into core particles and conferred protection against *B. pertussis* challenge in the Kendrick test, when used in conjunction with FHA.

2. Construction of yeast expression vector pWYG7

The fusion protein was expressed in *Saccharomyces cerevisiae* from the multi-copy vector pWYG7 which carried the regulated GAL7 promoter. The vector pWYG7, constructed at Wellcome, was used for the expression of HBP, i.e. of a fusion protein composed of a major epitope of P.69 fused to the amino-terminus of HBcAg. Vector pWYG7 is derived from the 2 μ vector pJDB219 (Beggs, Nature 275, 104-109, 1978) modified to contain a kanamycin-resistance mark r (kan^r) and the yeast galactose-regulated GAL7 promoter.

The construction of pWYG7 is outlined in Fig. 1. First the kan^r marker (HincII fragment from pUC4K; Vieira and Messing, Gene 19, 259, 1982) was ligated into the unique SmaI site of pJDB219 to give the kan^r tet^r vector pJDB219K. Secondly, a synthetic GAL7 promoter fragment (XhoI-BamHI fragment, sequence

shown in Fig. 2) was cloned into the unique Sall and BamHI sites of pJDB219K. The resulting vector, pWYG7, has the GAL7 promoter with unique BamHI and BclII sites upstream of the yeast 2 μ plasmid FLP gene transcriptional terminator (Sutton and Broach, Mol. Cell. Biol. 5, 2770-2780, 1985). Foreign genes to be expressed from pWYG7 are inserted between the BamHI and BclII sites. The design of the GAL7 promoter fragment is discussed below.

The smallest fragment of DNA upstream of the GAL7 gene which exhibits full promoter activity has been defined by deletion mapping (Tajima *et al.*, Yeast 1, 67-77, 1985). Based on this information a 260 bp GAL7 promoter fragment was synthesised (Fig. 2 for sequence). The 260bp promoter was synthesised as four overlapping oligonucleotides using a Pharmacia Gene Assembler (protocol supplied by Pharmacia). These oligonucleotides were phosphorylated and annealed using standard techniques, then ligated into XhoI-BamHI cut pIC-20H (Marsh *et al.*, Gene 32, 481-485, 1984). Positive clones were identified and their DNA sequenced using the double-stranded DNA sequencing method with universal and reverse sequencing primers (Hong, Biosci. Report 2, 907, 1982). The sequence of the GAL7 inserts was confirmed, and then the XhoI-BamHI GAL7 insert was excised and cloned into pJDB219K as described above.

The design of the GAL7 promoter fragment in pWYG7 is such that the natural GAL7 DNA sequence has been slightly modified (2bp changed) in order to make the BamHI cloning site upstream of the GAL7 mRNA start sites. The foreign gene to be expressed is then linked with synthetic DNA to the BamHI site, such that the GAL7 mRNA start sites are introduced, along with the GAL7 upstream untranslated sequences. Thus the first non-yeast DNA downstream of the promoter is the initiating ATG codon of the foreign gene, and the transcript produced will have a yeast GAL7 leader rather than a foreign leader which could reduce efficiency of translation.

3. Construction of the HBP fusion protein yeast expression vector pWYG7HBP

A BamHI-BamHI expression unit for HBP was assembled in an intermediate vector, pKGC-69K. This vector, which contains GAL7 sequences upstream of the initiator ATG codon, was made by first constructing a HBcAg vector, pKGC, with an NcoI (CCATGG) site engineered at the initiator ATG. The DNA encoding the predicted B.pertussis epitope was then inserted into this site as a synthetic NcoI-NcoI oligonucleotide linker. The overall scheme for the construction of pKGC, pKGC-69K, and the final expression vector, pWYG7HBP, is shown in Fig. 3.

(i) Construction of pKGC

A synthetic Sall-EcoRI linker containing the 3' terminal sequences of HBcAg (Aval site to stop codon) was ligated between the Sall and EcoRI sites of pUC18 (Vieira and Messing, 1982) to give the plasmid pKGF. The rest of the HBcAg gene was then assembled in pKGF in a three-way ligation: the 2.7kb HindIII-Aval fragment of pKGF was isolated and ligated to (i) an 87bp synthetic (phosphorylated) HindIII-MaeIII fragment (oligoA, Fig. 4a) containing yeast GAL7 upstream sequences and the 5' end of the HBcAg gene, and (ii) a 500 bp MaeIII-Aval fragment from pEB208 (Clarke *et al.*, Nature 330, 381-384, 1987), which contains most of the central part of the gene. The resulting plasmid, pKGC, is an intermediate vector for expression of HBcAg from pWYG7. The sequence of the synthesized regions of pKGC was confirmed using the double-stranded method with universal and reverse sequencing primers (Hong, 1982).

(ii) Construction of pKGC-69K

The HBcAg gene in pKGC had been modified to have a unique NcoI (CCATGG) site at the initiator ATG. Therefore N-terminal peptide fusions could be made by inserting suitable NcoI-NcoI DNA fragments at this site. The epitope of the P.69 antigen was predicted to occur at the Pro-rich repeat region, so a suitable pair of oligonucleotides were synthesised to encode this epitope. The oligonucleotides made and the corresponding amino acid sequence are given in Fig. 4b; the B. pertussis DNA was altered to give a yeast-optimal codons. pKGC was digested with NcoI, and the annealed, unphosphorylated oligonucleotides were cloned in using the linker-tailing method (Lathe *et al.*, BRL Focus 6, issue 4, 1984). From the resulting plasmids with inserts, ones of the required orientation were selected by double-stranded sequencing of the inserts. The resulting vector, pKGC-69K could be used as a source of a BamHI-BamHI fragment for transfer to pWYG7.

(iii) Construction of pWYG7HBP

The BamHI-BamHI fragment from pKGC-69K was isolated and cloned into pWYG7 (*dam*⁻ DNA) that had been digested with BamHI and BclI and then treated with calf-intestinal alkaline phosphatase. After transformation, *Kan*^r colonies were tested for inserts of the correct orientation, and these plasmids were denoted pWYG7HBP. *E. coli* MC1061 containing pWYG7HBP was deposited on 28 July 1989 at the National Collection of Industrial and Marine Bacteria (NCIMB), Aberdeen, GB under accession number NCIMB 40176.

4. Transformation of yeast with pWYG7HBP

The vector pWYG7HBP was introduced into the *Saccharomyces cerevisiae* strain S150-2B (a, *leu2*, *his3*, *ura3*, *trp1*, McCleod et al., Cold Spring Harbor Symp. Quant. Biol. 49, 779-787, 1984) using the lithium transformation procedure of Ito et al. (J. Bact. 153, 163-168, 1983). Transformed yeast cells were incubated in YPD broth (Sherman et al., Methods in Yeast Genetics, Cold Spring Harbor, New York, 1983) at 30°C overnight prior to plating out on selective medium (YPD plus 500 µg/ml G418). This allows expression of G418-resistance and increases transformation frequency. Colonies that came up as G418^r were checked on minimal medium lacking leucine (YNB + glucose + histidine + uracil + tryptophan, Sherman et al., 1983) to check for the *Leu*⁺ phenotype also conferred by pWYG7HBP. Positive transformants (G418^r *Leu*⁺) were used for expression analysis.

5. Galactose-induction of expression of HBP

Transformants were grown to the mid-logarithmic stage (10⁷ cells/ml) in YP broth containing 2% raffinose and 500 µg/ml G418 at 30°C in an orbital shaker. An aliquot of 40% galactose was then added to a final concentration of 2%, and the culture was incubated for a further 48h. The cells were then harvested by low speed centrifugation, washed once in distilled water, and resuspended in ice-cold break buffer (20mM sodium phosphate pH7.0, 0.1% Triton X-100, 4mM phenylmethanesulphonyl fluoride, 4mM EGTA, and 2µg/ml each of pepstatin, antipain, leupeptin, and chymostatin; 5ml for cells from a 250ml culture). Acid-washed glass beads (0.45mm) were added and the cells were broken by vigorous vortexing. The crude cell lysate was cleared by centrifugation for 15 mins at 10,000g. The protein concentration of the cleared supernatant was determined using the BioRad protein assay (BioRad, according to manufacturer's instructions), and the material was stored at -20°C in aliquots.

6. Analysis of cell lysates for expression

The proteins in the induced yeast cell lysates were analysed by separation in SDS-polyacrylamide gels (Laemmli, Nature 227, 680-685, 1970). 50 µg of soluble protein were loaded per track, and as a negative control an extract of induced S150-2B was loaded. A new protein band was detected by staining with Coomassie blue in the pWYG7HBP-transformed cell extracts, migrating at about 24,000kDa (not shown). It was confirmed that this polypeptide is HBP by using Western blot analysis with either HBcAg or BB05 antiserum (Fig. 5, result with HBcAg serum). ELISA quantitation data indicate expression levels of 10-30% of cell protein.

7. HBP fusion protein expressed in yeast associates to form core particles

In order to test whether or not the HBP protein made in yeast was present as core particles, the induced cell extract was layered over a 15%-45% w/v sucrose density gradient (in phosphate-buffered saline) and centrifuged in a Beckman SW28 rotor (28,000rpm, 4h). The gradient was fractionated and the fractions analysed for the presence of HBcAg- or *B. pertussis* epitope-reacting material by spotting aliquots from each gradient fraction onto a nitrocellulose grid and processing the filter as for Western blot (dot blot shown in Fig. 6).

Peak reactivity was detected in the middle fractions of the gradient, and none was detected at the top of the gradient. This indicates that all the HBP protein made in yeast associates to form core particles which sediment in a sucrose gradient. For confirmation, an aliquot from a peak gradient fraction was sent for electron microscopy (phosphotungstic acid stain). A large number of viral core particles were clearly seen (not shown).

Example 3: Dot-blot antibody hybridisation

Aqueous solutions of the three peptides prepared in Example 1 (2mg/ml) and the HBP fusion protein prepared in Example 2 were used in dot-blot antibody hybridization experiments. Briefly, 10µl samples of the peptide or fusion protein were applied to a nitrocellulose membrane and allowed to dry on. Filter hybridization experiments with monoclonal antibodies were carried out by normal methods (see, for example, "Antibodies, a laboratory manual" p.178, Ed. E. Marlow and D. Lane 1988, Cold Spring Harbor Laboratory). The Mabs were:

BB05 and BB07 (Montarez *et al.*, Infect. Immun. 47, 744, 1985) which were raised against the 68kDa protein from *B. bronchiseptica* but which cross-react with P.69 from *B. pertussis*; and a panel of 7 Mabs raised against *B. pertussis*.

The results are shown in Table 1 and Figure 7. Figure 7 shows the region of P.69 around which the peptides were made. The three chemically synthesised peptides have an additional non-natural carboxy-terminal cysteine residue. The HBP fusion protein has a N-terminal methionine residue. The results establish that the epitope of interest lies within the region with the following sequence:

APQPGPQPPQPPQPQPEAPAPQP.

Table 1

	Monoclonal antibody	Peptide			
		683	684	685	HBP fusion protein
<u><i>Bordetella bronchiseptica</i></u>	BB05	+	-	-	+
	BB07	+	-	-	-
<u><i>Bordetella pertussis</i></u>	BPE3	-	-	+	-
	BPD8	-	-	-	-
	BPE8	-	-	-	-
	D5E9	-	-	-	-
	F6E5	-	-	-	-
	E4A8	+	-	-	-
	E4D7	+	-	-	-

Example 4: Kendrick Test1. Coupling of peptides

The peptides of Example 1 were coupled to Keyhole Limpet Haemocyanin (KLH) through an added C-terminal cysteine residue using M-Maleimidobenzoyl-N-hydroxysulfosuccinimide ester (MBS) as a heterobifunctional cross-linker (Liu, F. *et al.* Biochemistry 18, 690, 1979). Briefly, dialysed KLH is reacted with MBS dissolved in dimethylformamide (DMF) at room temperature, then run through a G-25 column. The KLH protein peaks were pooled, and added to free peptide. The pH was then adjusted to between 7 and 7.5, and after 3 hours stirring at room temperature the coupled peptide was stored at -20°C. The KLH concentration must not exceed 20mg/ml, MBS/KLH molar ratio is 40:1, and the final concentration of DMF in the reaction should be below 30%. Peptides 683, 684 and 685 were coupled using 2.5mg KLH per 5mg peptide (a ratio of 1:2 wt/wt).

2. Preparation of Filamentous Haemagglutinin (FHA)

FHA can be prepared by methods well known in the art (see Arach and Munoz J.J. (1970), Infect. Immunology 25 764-767; Ashworth *et al.* (1982) Infect. Immun. 37 1278-1281). However FHA in the following procedure was prepared in accordance with the following protocol.

FHA purification: *B. pertussis* Tomaha or BP357 (Tn5 transposon mutant of A.A. Weiss *et al.* (1983) which does not secrete the LPF) were grown in Stainer & Scholte medium (0.05 Tris) in 650 ml Costar flasks (150 ml in each) for 5 days at 37°C (Sato *et al.*, Infection and Immunity 41, 313-320, 1983). Before centrifugation (30 mins at 6000 x g) 50µM 1,10-phenanthroline monohydrate as proteolysis inhibitor was

added to the cultures. The cell free supernatant was applied to a 30 x 150 mm column of Hydroxylapatite (BD4) and washed (all at room temperature and a speed of pumping 500 ml/hr) sequentially with 10mM phosphate buffer, pH 8, 100 mM phosphate buffer, pH 7.1 until stable baseline.

The retained material was eluted with 0.5M NaCl added to the 100mM phosphate buffer and the peak fractions agglutinating goose red blood cells were pooled. The pool was dialyzed overnight against 25-30 volumes of 0.025 M Bis-Tris/HCl buffer at 4°C. The precipitated FHA was collected on a centrifuge (20 mins at 8000 x g). The next step was inspired by Cowell *et al* (in vol. IV, Bacterial Vaccine, 371-379, Seminars in Infectious Disease, Weinstein and Fields: editors, Thieme Verlag, New York, Stuttgart, 1982) who found that the FHA (as well as LPF) is soluble in 40mM beta-alanine buffer, pH 3.5. The precipitated FHA was solubilized in the smallest possible volume of 3-alanine buffer (3.57g 3-alanine and 0.35g formic acid per litre), insolubles removed by centrifugation and the clear supernatant was applied to a column (25 x 500mm) of Ultrogel AcA 34 equilibrated and eluted with the same buffer.

The haemagglutinating material appeared in a sharp peak followed by a shoulder. The shoulder material was discarded, the fractions from the sharp peak were pooled and kept frozen or re-precipitated by dialysis against 0.025M Bis-Tris buffer and dissolved in a smaller volume of 3-alanine buffer. The solubility is approx. 2.5 mg FHA/ml. The material frozen (at -20 or -40°C) at acid pH appeared stable as judged from its ELISA reactivity and appearance in the SDS-PAGE gels: it formed prevalently three strong bands in the region of 150-100KD.

20 3. Kendrick Test

This was performed according to W.H.O. Requirements for Pertussis Vaccine using MF1 or NIH Mice (OLAC, category 3, free of most pathogens including *B. brochi*septica), weighing 14-16g. The antigens, in 0.5ml volumes, were inoculated concomitantly intraperitoneally and comprised a top dilution and three four-fold serial dilutions. After two weeks the mice were challenged intracerebrally using the recommended challenge strain 18-323 (100-200 LD₅₀). The number of survivors in each group was used for calculation of the relative potency in respect to the British Pertussis Reference Vaccine 66/84 using a program of parallel line probit analysis. The results are shown in Table 2.

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Table 2Vaccine

5

	Peptides (μ g)	683	684	685	+FHA(μ g)	Survivors
10	1	10	10	10	20	6/16
	2	3.3	3.3	3.3	6.6	3/16
15	3	1.11	1.11	1.11	2.2	7/16
	4	0.37	0.37	0.37	0.74	5/16

20

HBP fusion Protein (μ g) + FHA(μ g)

	1	1	20	8/16
	2	0.33	6.6	11/16
25	3	0.11	2.2	4/16
	4	0.037	0.74	1/16

30

FHA alone (μ g)

	1	20	3/16
35	2	6.6	2/16
	3	2.2	3/16
40	4	0.74	1/16

45

66/88 whole cell vaccine I.U. Survivors

	1	0.25	11/16
	2	0.08	2/16
50	3	0.028	2/16
	4	0.009	3/16

The results show clearly that the combination of FHA with either a mixture of all three chemically synthesized peptides or with the HBP fusion protein is more potent than FHA alone.

EXAMPLE 5: Epitope mapping using synthetic peptides(Pepscan)

Hexameric peptides, overlapping by one amino acid residue, were synthesised on solid phase polyethylene pins as described by Geysen *et al* (PNAS USA 81, 1984, 3998-4002). Ninety four hexameric peptides were synthesised, covering residues 505 to 603 of the P.69 antigen of *B. pertussis*. Pepscan peptide 1 was Thr(505)-Asp(510). Pepscan peptide 94 was Ala(598)-Leu(603).

Reactivity of the peptides to monoclonal antibody (mAb) BB05 was determined by incubation of the pins in antibody for 1-2 hrs or overnight, followed by washing and incubation in peroxidase conjugated goat-antimouse antibody. The mAb BB05 is an (IgG1) raised against the P.68 antigen from *B. bronchiseptica* - (Montarex *et al*, Infect. Immun. 47, 1985, 644-751). This mAb is a neutralizing mAb which cross-reacts with P.69 from *B. pertussis*.

Enzyme reactivity was determined by incubation of the pins in ABTS substrate solution (50 mg azinod-3-ethyl-benzthiazodinsulphonate (ABTS) in 100ml buffer (0.1 molar disodium hydrogen orthophosphate, 0.08 molar citric acid, pH 4.0 containing 30µl hydrogen peroxide)), and the A_{420} of the solution measured after 10-60 minutes using a Titertek Multiscan MC1100. Pins were sonicated for 30 min at 65°C in 0.1M Na_2HPO_4 , 0.1%SDS, 0.01 M B-mercaptoethanol to remove bound antibody and stain complexes prior to incubation in subsequent antibody. The results are shown in Figure 8.

As can be seen from Figure 8, mAb BB05 recognised only Pepscan peptide 43 (sequence PGPQPP). This is consistent with recognition of peptide 683 and located the BB05 epitope in the region of the (Pro-Gln-Pro)₅ repeats of P.69.

Claims

1. A polypeptide which presents an epitope comprising:
 - (a) the amino acid sequence coded for by nucleotides CCGGGTCCCCAGCCGCCG which are nucleotides 1885 to 1902 of the P.69 gene of *B.pertussis* CN2992; or
 - (b) the corresponding amino acid sequence of another strain of *B.pertussis* or of a strain of *B.parapertussis* or *B.bronchiseptica*;
 the said polypeptide being no more than 50 amino acid residues long or being a chimaeric protein having an amino acid sequence comprising the sequence of a carrier protein and a foreign sequence of no more than 50 amino acid residues which comprises the sequence of a said epitope.
2. A polypeptide according to claim 1, wherein the said epitope comprises
 - (a₁) the amino acid sequence coded for by nucleotides

CGGCCGAGCCGGGTCCCCAGCCGCCGAGCCGCCGAGCCGCCGAGCCGCCGAGC

CGGAAGCGCCGGCGCCGCAACCG

which are nucleotides 1876 to 1944 of the P.69 gene of *B.pertussis* CN2992; or

(b₁) the corresponding amino acid sequence of another strain of *B.pertussis* or of a strain of *B.parapertussis* or *B.bronchiseptica*;

3. A polypeptide according to claim 1 or 2, which is composed of no more than 50 amino acid residues and which is provided with a cysteine residue at the N-terminus and/or the C-terminus.
4. A polypeptide according to any one of the preceding claims, which is composed of up to 30 amino acid residues.
5. A polypeptide according to claim 1 or 2, which is a fusion protein comprising hepatitis B core antigen to the amino terminus of which is linked the said foreign sequence.
6. A polypeptide according to claim 1, which has the amino acid sequence:
APQPGPQPQPQPQPEAPAPQPPAGRELSC or
PGPQPP;

or which has the amino acid sequence:

MAPPAPKPAPQPGPQPPQPPQPQPEAPAPQP

fused to the amino terminus of hepatitis B core antigen.

- 5 7. A process for the preparation of a polypeptide as defined in claim 1, which process comprises condensing single amino acids and/or preformed peptides of two or more amino acids in the order in which amino acids occur in the said polypeptide.
8. A process for the preparation of a polypeptide as defined in claim 1, which process comprises:
 - 10 (i) preparing an expression vector which incorporates a DNA sequence encoding the said polypeptide and which is capable of expressing the said polypeptide when provided in a suitable host; and
 - (ii) providing the said vector in the said host such as to enable expression of the said polypeptide to occur.
- 15 9. A conjugate comprising a polypeptide as defined in claim 1 linked to a physiologically acceptable carrier.
10. A vaccine comprising a pharmaceutically acceptable carrier or diluent, a polypeptide as defined in claim 1 and, optionally, filamentous haemagglutinin.

Patentansprüche

1. Polypeptid, welches ein Epitop darstellt, umfassend:
 - 25 (a) die durch die Nucleotide CCGGGTCCCCAGCCGCCG, welche die Nucleotide von 1885 bis 1902 des P.69-Gens von *B. pertussis* CN2992 sind, codierte Aminosäuresequenz; oder
 - (b) die entsprechende Aminosäuresequenz eines anderen Stammes von *B. pertussis* oder eines Stammes von *B. parapertussis* oder *B. bronchiseptica*;
 wobei das Polypeptid nicht mehr als 50 Aminosäurereste lang ist oder ein chimäres Protein mit einer Aminosäuresequenz ist, die die Sequenz eines Trägerproteins und einer Fremdsequenz von nicht mehr

30 als 50 Aminosäureresten einschließt, welches die Sequenz des Epitops enthält.
2. Polypeptid nach Anspruch 1, wobei das Epitop folgendes umfaßt:
 - (a₁) die durch die Nucleotide

GCGCCGCAGCCGGGTCCCCAGCCGCCG-
CAGCCGCCGCAGCCGCAGCCGGAAGCGCCGGCGCCGCAACCG,

- 40 welche die Nucleotide von 1876 bis 1944 des P.69-Gens von *B. pertussis* CN2992 sind, codierte Aminosäuresequenz; oder
- (b₁) die entsprechende Aminosäuresequenz eines anderen Stammes von *B. pertussis* oder eines Stammes von *B. parapertussis* oder *B. bronchiseptica*.
- 45 3. Polypeptid nach Anspruch 1 oder 2, welches aus nicht mehr als 50 Aminosäureresten besteht, und welches mit einem Cysteinrest am N-Terminus und/oder C-Terminus versehen ist.
4. Polypeptid nach mindestens einem der vorhergehenden Ansprüche, welches aus bis zu 30 Aminosäureresten besteht.
- 50 5. Polypeptid nach Anspruch 1 oder 2, welches ein Fusionsprotein ist, das ein Hepatitis-B-Kern-Antigen am Amino-Terminus, an das die Fremdsequenz angehängt ist, umfaßt.
6. Polypeptid nach Anspruch 1, welches die Aminosäuresequenz

65 APQPGPQPPQPPQPQPEAPAPQPPAGRELSC

oder PGPQPP aufweist;

oder welches die Aminosäuresequenz

MAPPAPKPAPQPGPQPPQPPQPQPEAPAPQP,

fusioniert am Amino-Terminus des Hepatitis-B-Kern-Antigens, enthält.

7. Verfahren zur Herstellung eines Polypeptids nach Anspruch 1, wobei das Verfahren das Kondensieren von einzelnen Aminosäuren und/oder vorgebildeten Peptiden aus zwei oder mehreren Aminosäuren in der Reihenfolge umfaßt, in der Aminosäuren in dem Polypeptid auftreten.
8. Verfahren zur Herstellung eines Polypeptids nach Anspruch 1, wobei das Verfahren folgendes umfaßt:
 - (i) das Herstellen eines Expressionsvektors, welcher eine für das Polypeptid codierende DNA-Sequenz enthält und welcher zur Expression des Polypeptids in der Lage ist, wenn er in einen geeigneten Wirt eingeführt ist; und
 - (ii) das Bereitstellen des Vektors in dem Wirt dergestalt, daß die Expression des Polypeptids ermöglicht wird.
9. Konjugat, umfassend ein Polypeptid nach Anspruch 1, das an einem physiologisch annehmbaren Träger gebunden ist.
10. Impfstoff, umfassend einen pharmazeutisch annehmbaren Träger oder ein pharmazeutisch annehmbares Verdünnungsmittel, ein Polypeptid nach Anspruch 1 und gegebenenfalls filamentöses Hämagglutinin.

Revendications

1. Polypeptide qui présente un épitope comprenant :
 - (a) la séquence d'acides aminés codées par les nucléotides CCGGGTCCCCAGCCGCCG qui sont les nucléotides 1885 à 1902 du gène P.69 de B.pertussis CN2992 ; ou
 - (b) la séquence d'acides aminés correspondante d'une autre souche de B.pertussis ou d'une souche de B.parapertussis ou B.bronchiseptica ;
 ledit polypeptide ne comprenant pas plus de 50 résidus d'acides aminés ou étant une protéine chimère ayant une séquence d'acides aminés comprenant la séquence d'une protéine porteuse et une séquence étrangère, constituée au plus de 50 résidus d'acides aminés, qui comprend la séquence dudit épitope.

2. Polypeptide selon la revendication 1, dans lequel ledit épitope comprend
 - (a₁) la séquence d'acides aminés codées par les nucléotides

GCGCCGCAGCCGGGTCCCCAGCCGCCGCAGCCGCCGCAGCCGCAGCCGGAAGCGCCGGC
GCCGCAACCG

- qui sont les nucléotides 1876 à 1944 du gène P.69 de B.pertussis CN2992 ; ou
- (b₁) la séquence d'acides aminés correspondante d'une autre souche de B.pertussis ou d'une souche de B.parapertussis ou B.bronchiseptica ;

3. Polypeptide selon la revendication 1 ou 2, qui est constitué au plus de 50 résidus d'acides aminés et qui comprend un résidu cystéine à l'extrémité N-terminale et/ou l'extrémité C-terminale.
4. Polypeptide selon l'une quelconque des revendications précédentes qui est composé d'un maximum de 30 résidus d'acides aminés.
5. Polypeptide selon la revendication 1 ou 2, qui est une protéine de fusion comprenant l'antigène core de l'hépatite B, à l'extrémité aminoterminal duquel est liée ladite séquence étrangère.
6. Polypeptide selon la revendication 1, qui a la séquence d'acides aminés :

APQPGPQPPQPPQPEAPAPQPPAGRELSC ou
PGPQPP ;

 ou qui a la séquence d'acides aminés :

MAPPAPKPAPQPGPQPPQPPQPEAPAPQP

fusionnée à l'extrémité aminoterminal de l'antigène core de l'hépatite B.

- 5
7. Procédé de préparation d'un polypeptide tel que défini dans la revendication 1, procédé qui comprend la condensation d'acides aminés isolés et/ou de peptides préformés de deux acides aminés ou plus, dans l'ordre dans lequel les acides aminés se présentent dans ledit polypeptide.
8. Procédé de préparation d'un polypeptide tel que défini dans la revendication 1, procédé qui comprend les étapes consistant à :
- 10 (i) préparer un vecteur d'expression qui contient une séquence d'ADN codant pour ledit polypeptide et qui est capable d'exprimer ledit polypeptide lorsqu'il est introduit dans un hôte approprié ; et
(ii) introduire ledit vecteur dans ledit hôte, afin de permettre l'expression dudit polypeptide .
9. Conjugué comprenant un polypeptide tel que défini dans la revendication 1, lié à un porteur physiologiquement acceptable.
- 15
10. Vaccin comprenant un véhicule ou un diluant pharmaceutiquement acceptable, un polypeptide tel que défini dans la revendication 1 et, éventuellement, de l'hémagglutinine filamenteuse.
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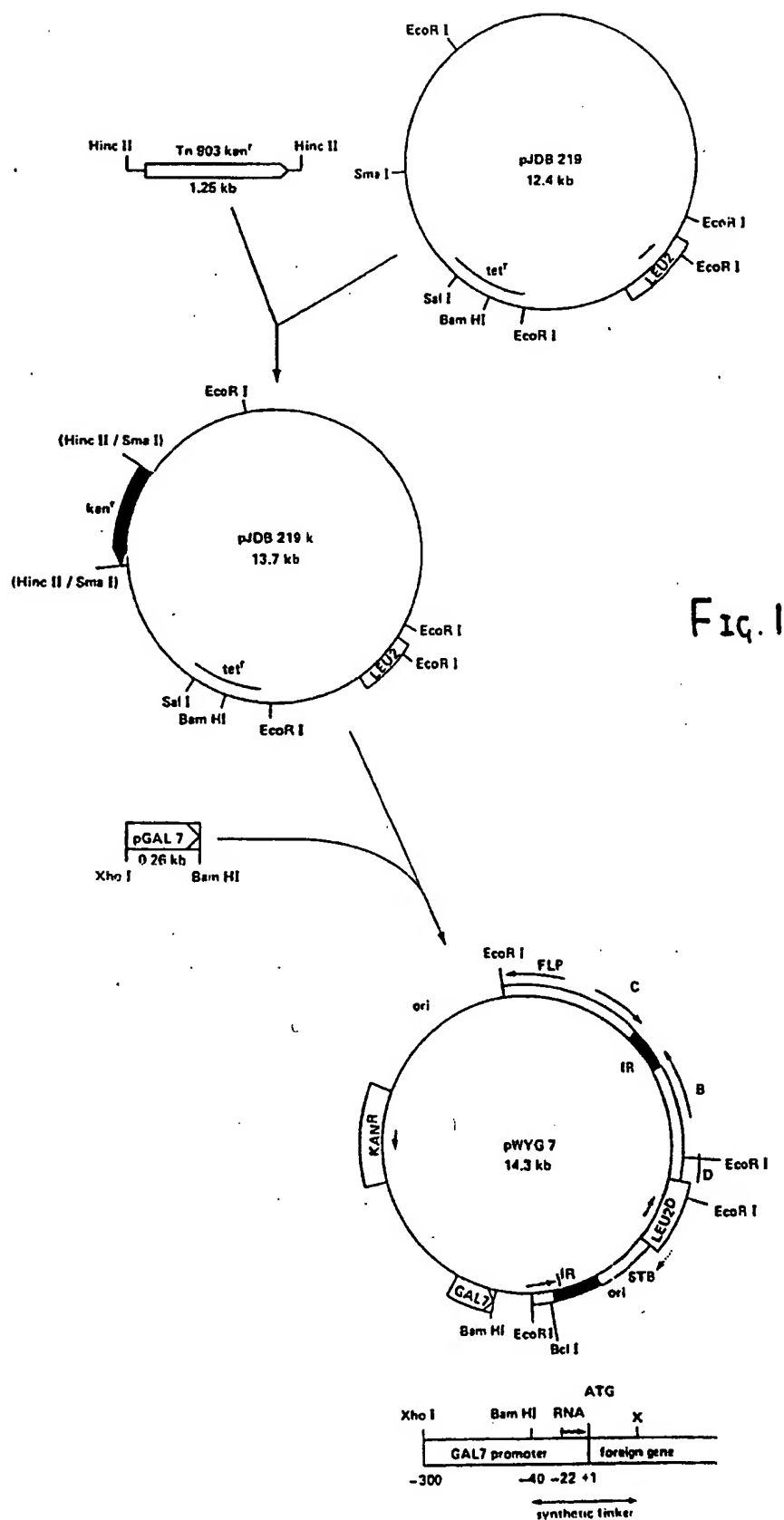


FIG. 1

```

      10      20      30      40      50      60
XhoI  .  .  .  .  .  .  .  .  .  .  .  .
CTCGAGACGT CTATACTTCG GAGCACTGTT GAGCGAAGGC TCATTAGATA TATTTTCTGT
GAGCTCTGCA GATATGAAGC CTCGTGACAA CTCGCTTCCG AGTAATCTAT ATAAAAGACA

      70      80      90     100     110     120
.  .  .  .  .  .  .  .  .  .  .  .
CATTTTCCTT AACCCAAAAA TAAGGGAGAG GGTCCAAAAA GCGCTCGGAC AACTGTTGAC
GTAAAAGGAA TTGGGTTTTT ATTCCCTTTC CCAGGTTTTT CGCGAGCCTG TTGACAACTG

      130     140     150     160     170     180
.  .  .  .  .  .  .  .  .  .  .  .
CGTGATCCGA AGCACTGGCT ATACAGTGTT CACAAAATAG CCAAGCTGAA AATAATGTGT
GCACTAGGCT TCCTGACCGA TATGTCACAA GTGTTTTATC GGTTCGACTT TTATTACACA

      190     200     210     220     230     240
.  .  .  .  .  .  .  .  .  .  .  .
AGCCTTTAGC TATGTTTCAGT TAGTTTGGCT AGCAAAGATA TAAAAGCAGG TCGGAAATAT
TCGGAAATCG ATACAAGTCA ATCAAACCGA TCGTTTCTAT ATTTTCGTCC AGCCTTTATA

      250     260     270     280     290     300
.  .  .  .  .  .  .  .  .  .  .  .
TTATGGGCAT TATTATGCAG AGGATCCACA TGATAAAAAA AACAGTTGAA TATCCCTCA
AATACCCGTA ATAATACGTC TCCTAGGTGT ACTATTTTTT TTGTCAACTT ATAAGGGAGT

      310
.  .  .  .  .  .  .  .  .  .  .  .
AAAATGACTG
TTTACTGAC

```

Fig. 2

Fig. 3.

(a)

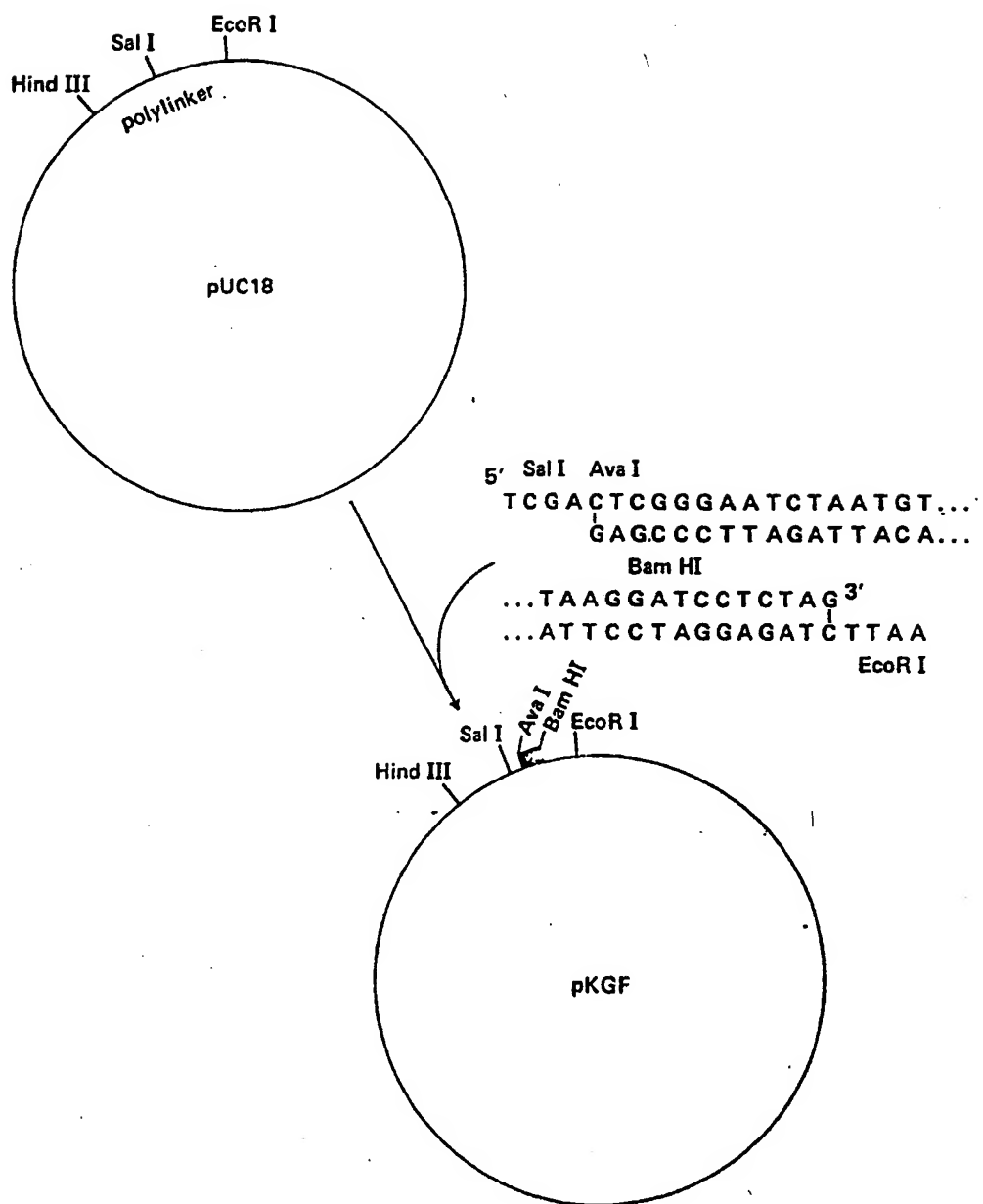


Fig. 3.

(b)

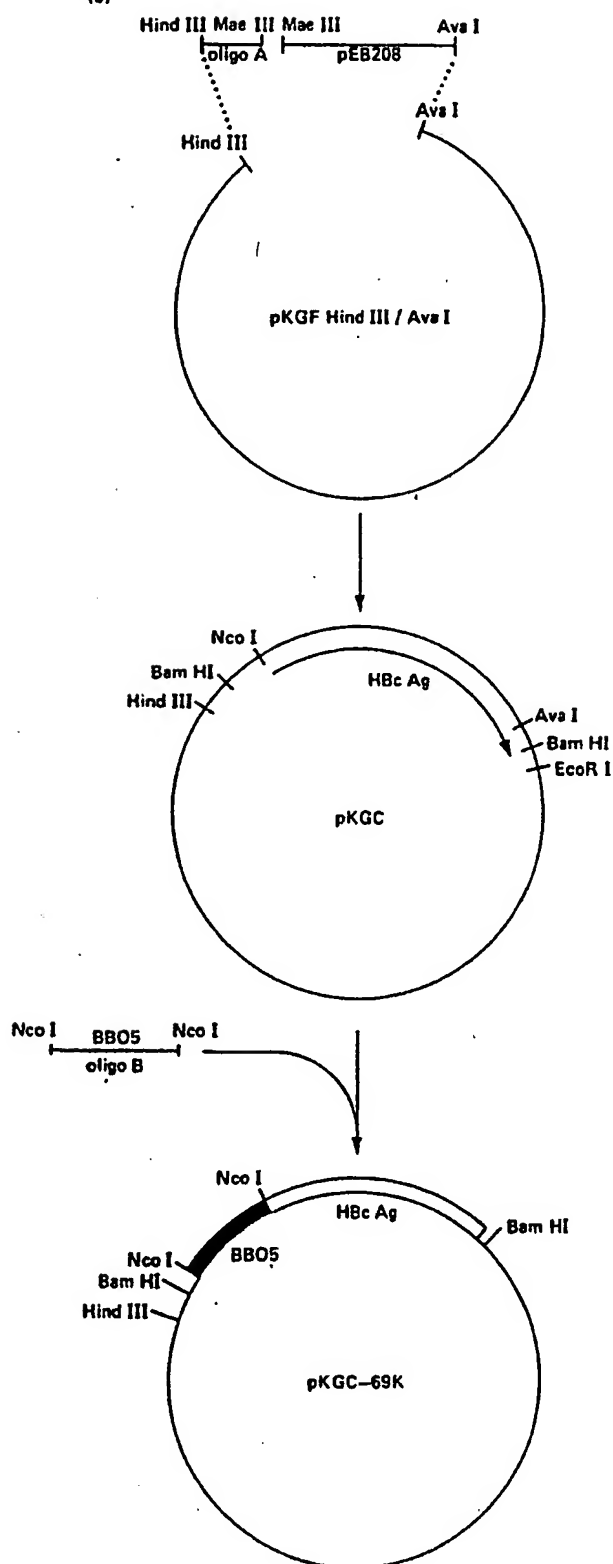


Fig. 3.

(c)

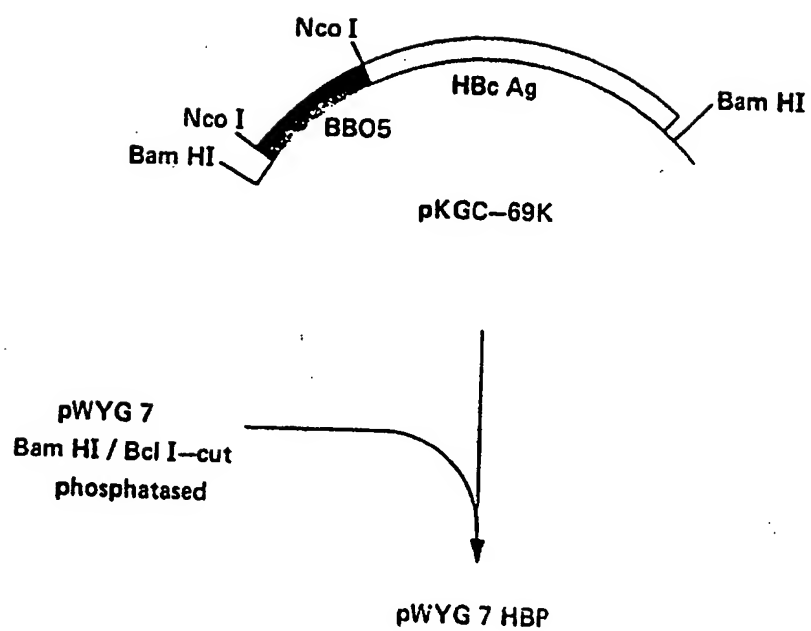


Fig. 4a.

Hind III Bam HI GAL7 leader Nco I

5' AGCTTGGATCCACATGATAAAAAAACAGTTGAATATTCCCTCAACCATGG...
 3' ACCTAGGTGTACTATTTTTTTTGTCAACTTATAAGGGAGTTGGTACC...

...ACATTGACCCTTATAAAGAATTCTGGAGCTACTGTGGA 3'
 ...TGTAAGTGGGAATATTTCTTAAGCCTCGATGACACCTCAATG 5'

HBcAg gene 5' end Mae III

Fig. 4b.

Nco I M A P P A P K P A P Q P G P Q P P

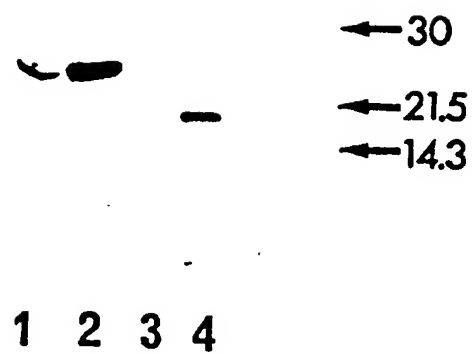
5' CATGGCTCCACCAGCTCCAAAACCAGCTCCACAACCAGGTCCACAACCACCA..
 3' CGAGGTGGTCTGAGGTTTTGGTCTGAGGTGTTGGTCCAGGTGTTGGTGGT..

Q P P Q P Q P E A P A P Q P

...CAACCACCACAACCACAACCAGAAGCTCCAGCTCCACAACC 3'
 ...GTTGGTGGTGGTGGTGGTCTTCGAGGTCTGAGGTGTTGGGTAC 5'

Nco I

Fig. 5



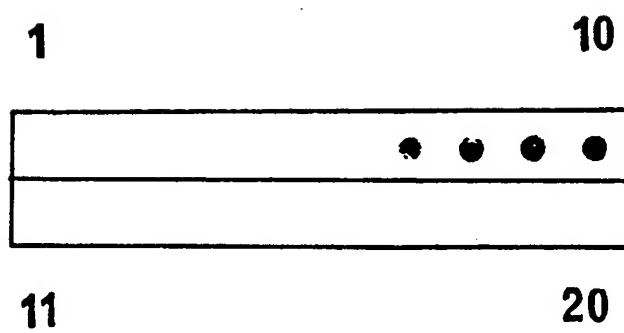
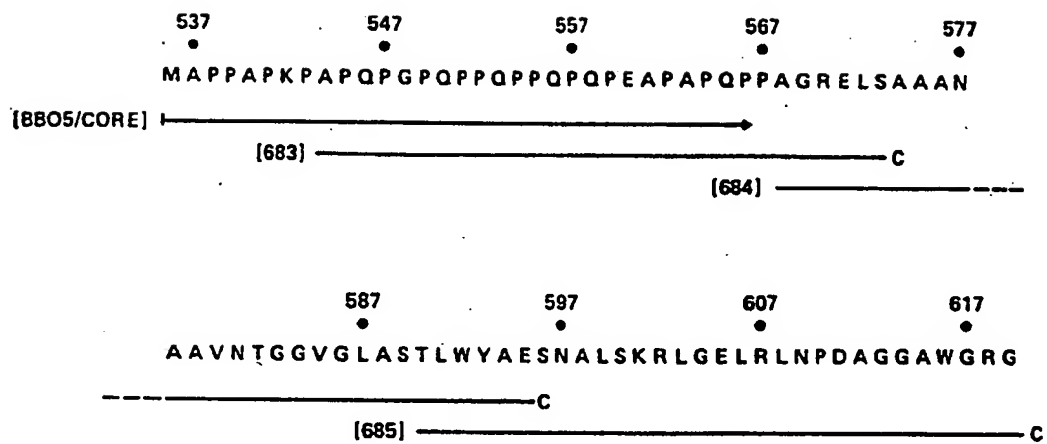


Fig. 6

Figure 7



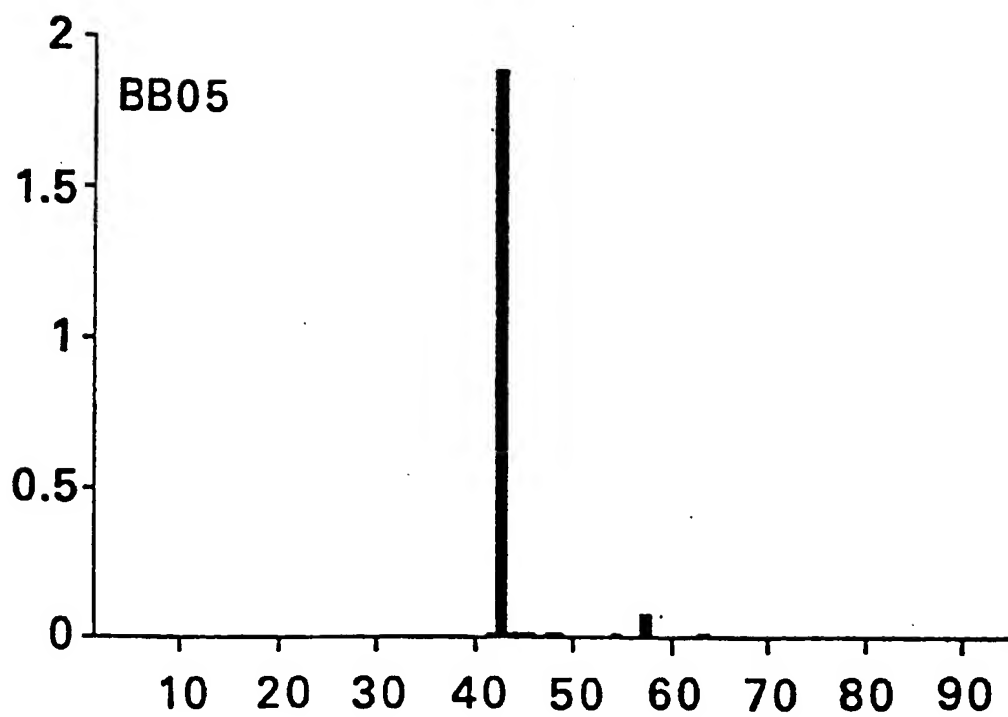


Fig. 8